



Compatibility of Fungal Root Endophyte *Piriformospora indica* with New Generation Fungicides

**Padhiyil Amrutha ^{a,b*}, Elizabeth T. Jojy ^a,
Joy Michal Johnson ^c, Sarada S. ^d, Sajeena A. ^e,
Radhakrishnan N. V. ^a and Swapna Alex ^f**

^a Department of Plant Pathology, College of Agriculture, Vellayani (Kerala Agricultural University) 695522, Thiruvananthapuram, Kerala, India.

^b Division of Plant Pathology, School of Agricultural Sciences, Karunya Institute of Technology and Sciences, Coimbatore, Tamil Nadu, India.

^c Coconut Research Station (KAU), Balaramapuram 695501, Thiruvananthapuram, Kerala, India.

^d Department of Vegetable Science, College of Agriculture, Vellayani, 695522, Thiruvananthapuram, Kerala, India.

^e Integrated Farming System Research Station, Karamana, Thiruvananthapuram, Kerala 695002, India.

^f Department of Plant Biotechnology, College of Agriculture, Vellayani, 695522, Thiruvananthapuram, Kerala, India.

Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

Article Information

DOI: <https://doi.org/10.9734/ijpss/2024/v36i64675>

Open Peer Review History:

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: <https://www.sdiarticle5.com/review-history/116370>

Original Research Article

Received: 22/02/2024

Accepted: 25/04/2024

Published: 19/05/2024

*Corresponding author: E-mail: amruthapvijayan8@gmail.com;

Cite as: Amrutha, P., Jojy, E. T., Johnson, J. M., Sarada S., Sajeena A., Radhakrishnan N. V., & Alex, S. (2024). Compatibility of Fungal Root Endophyte *Piriformospora indica* with New Generation Fungicides. *International Journal of Plant & Soil Science*, 36(6), 709–719. <https://doi.org/10.9734/ijpss/2024/v36i64675>

ABSTRACT

Colletotrichum gloeosporioides causing anthracnose is an important fungal disease of yard long bean infecting leaf, stem, petiole, flower and pod leading to significant yield loss. New generation fungicides viz. strobilurins and triazoles are widely used in the management of the disease. *Piriformospora indica* is a widely used beneficial root endophytic fungus that suppresses plant diseases in addition to enhanced growth promotion. The present study was outlined in completely randomized design (CRD) to test the compatibility of *P. indica* with the new generation fungicides commonly used in managing yard long bean anthracnose by poison food technique in petri dishes and broth media and by calculating the percentage of chlamydospore germinated. The results revealed that *P. indica* was compatible with strobilurins, combination fungicides of strobilurins and triazoles, carbendazim and pencycuron upto 90 per cent till 350 ppm in poison food and broth experiments. Moreover, germination of the chlamydospores was significant in number in these fungicides. But, triazole fungicides completely inhibited the mycelial growth and chlamydospore germination of *P. indica*. Thus, from the present experiment it was clear that *P. indica* is compatible with strobilurins and combination fungicides.

Keywords: *P. indica*; root endophyte; fungicides; compatibility; fungicide residue; dissipation.

1. INTRODUCTION

Yard long bean (*Vigna unguiculata* subsp. *sesquipedalis* (L) Verdcort) is a highly preferred protein rich vegetable with many health benefits. In Kerala it is grown for its long and succulent green pods. The crop is severely affected by fusarium wilt, viral diseases, anthracnose and cercospora leaf spot; among which, anthracnose caused by *Colletotrichum gloeosporioides* is one of the most important fungal disease [1], causing an yield loss of 50 per cent [2]. The disease is widely distributed in the Asian and African continents. On susceptible cultivars, typical anthracnose lesions (brown to tan, sunken, and lenticular) quickly grow and consolidate to girdle stems, peduncles, and petioles [3]. High susceptibility of the disease is due to the lack of genetic resistance, epidemiological conditions, cultivation practices and the emergence of fungicide-resistant strains of the pathogen. Management of the disease is currently carried out by spraying triazole fungicides and the combination fungicide carbendazim and mancozeb [4-7] and biocontrol agents like *Trichoderma* spp. [3].

Application of the fungicides against fungal infections is regarded as a common method to secure regular food supply [8]. According to preliminary statistics of the U.S. Geological Survey (USGS), dithiocarbamates, triazoles, strobilurins and its combination fungicides constitutes the major fungicide category among synthetic fungicides. These chemicals are highly efficient in action, but their rigorous use can damage the environment and pose severe health

hazards. In this context, the use of biocontrol agents is highly influential against plant diseases and can reduce chemical fungicide usage and its impending health hazards.

Endophytes, which inhabit the plant tissue, possess all the properties of biocontrol agents as they shield the plant throughout its life stages. They are now a viable substitute to agrochemicals. According to Cook and Baker [9], a mixed formulation of chemicals and biological control antagonists can extend the duration and activate disease control, which can reduce the quantity of chemical; residue buildup, lessen pathogen virulence and make it more prone to attack by antagonists [10]. *Piriformospora indica* is one such fungus that is axenically culturable, invades broad hosts, is root colonizable and is mycorrhiza like versatile endophytic fungus that helps in better plant growth and performance [11]. *P. indica* is a potent bioregulator as it enhances shoot and root proliferation by synthesizing phytohormones, a good biofertilizer that gives multifaceted responses against various plant diseases [12-18].

The compatible nature of strobilurin fungicides and carbendazim with *Trichoderma* spp., *Pseudomonas* Spp. and *Bacillus subtilis* was demonstrated by Sendhilvel et al. [19], Anand et al. [20], Archana et al. [21], Bagwan [22], Sarkar et al. [23], Ranganathswamy et al. ([24], whereas triazole fungicides were inhibitory to the bio-agents. It is our first effort to investigate the compatibility of strobilurin, triazole and its combination fungicides with *P. indica*. Henceforth, we attempted to find the compatible

new generation fungicides with *P. indica* under *in vitro* conditions under petri plate, broth and via chlamydospore germination.

2. MATERIALS AND METHODS

2.1 Maintenance and Multiplication of *P. indica*

P. indica (Accession No. INBA3202001787) was cultured in Potato Dextrose Agar (PDA) medium (pH-6.5). The culture was maintained at Department of Plant Pathology, College of Agriculture, Vellayani. Five mm mycelial discs were cut out from the actively growing regions of hyphae and placed at the middle of petri dishes. The plates were incubated at room temperature ($27\pm 1^\circ\text{C}$) and at 80 per cent humidity with 12 hour dark and light for 10 days. It was sub-cultured once in fifteen days for its maintenance [25]. Periodical re-isolation was carried out from the colonized roots to maintain the colonization efficiency of the endophyte. For further multiplication, *P. indica* was cultured in Potato Dextrose Broth (PDB) (pH-6.5) in conical flasks and incubated at 70 rpm for 21 days at 27°C to produce a sizable amount of mycelial mat.

2.2 Compatibility Assay

New generation systemic fungicides *viz.*, three strobilurin (trifloxystrobin 50 WG, kresoxymethyl 44.3 SC, azoxystrobin 23Sc), four triazole (hexaconazole 5 EC, difenoconazole 25% EC, propiconazole 25% EC, tebuconazole 25.9% EC), two combination (azoxystrobin 11% + tebuconazole 18.3% SC, trifloxystrobin 25% + tebuconazole 18.3% SC) fungicides, a contact fungicide (pencycuron 22.9% SC) and systemic fungicide (carbendazim 50WP – positive control) at 100, 250, 350, 500 and 1000 ppm concentrations were tested *in vitro* against *P. indica* by recording the radial mycelial growth and nature of mycelial growth at various time intervals.

Double strength PDA (50ml) and sterile distilled water (50ml) was prepared in 250ml conical flasks and sterilized. *In vitro* compatibility of *P. indica* with the fungicides was determined by preparing desired concentrations of chemicals in sterile water and dispersing it with molten double strength PDA aseptically. *P. indica* was cultured in poisoned-PDA plates and a mycelial plug of 5mm diameter was placed at the centre of the Petri plates. PDA medium without fungicide served as control. Each treatment was replicated

thrice. Observations of mycelial growth were recorded after 1, 3, 5, 7, 10 and 15 days after incubation along with the days taken for full growth of *P. indica*. The per cent inhibition of *P. indica* by each fungicide treatment was calculated using the formula suggested by Vincent [26].

$$\text{Per cent inhibition of growth} = \frac{C-T}{C} \times 100$$

C= Growth of *P. indica* in control (mm)

T= Growth of *P. indica* in treatment (mm)

One hundred ml PDB and 100 ml sterile water was prepared in 250ml conical flasks and sterilized. 10000 ppm concentration of stock solution of each fungicides were prepared in 100 ml sterile water and the concentration was diluted to 100, 250, 350, 500 and 1000 ppm by pipetting required amount from stock to each of the PDB containing flasks. 5 mm disc of *P. indica* culture was added to each conical flasks having different concentration of the fungicides. Control flasks were also maintained. The wet and dry weight of *P. indica* mycelium was recorded at 21 days of growth along with the control.

2.3 Spore Germination Inhibition Study

P. indica was cultivated in 100ml of PDB for 21 days under room temperature at 70 rpm for ample chlamydospore production and the pores were mixed with sterile water. Stock solutions of the fungicides were prepared at concentrations 100, 250, 350, 500 and 1000 ppm. Sterilized cavity slide and cover slips were used in the study. Each cavity slide had 50 μl of each fungicide concentration and 50 μl of *P. indica* spore suspension (10^{-6} ml^{-1}), thoroughly mixed, and incubated at room temperature. Germination of spores was recorded and per cent inhibition was calculated for each fungicide at different intervals.

2.4 Statistics

“The Kerala Agricultural University's GRAPES, (General R-shiny Based Analysis Platform Empowered by Statistics; <https://www.kaugrapes.com/home>) a R-based analysis platform was used for the statistical analysis. Using Duncan's Multiple Range (DMRT) and one-way Analysis of Variance (ANOVA), the treatment means were compared with a probability of 0.05 per cent level of significance.

All data shown are mean \pm standard deviation (SD) of at least three biological replicates" [27].

3. RESULTS

3.1 Compatibility of Systemic Fungicides with *P. indica* under *in vitro*

A minimal inhibition of 1.12 per cent was observed with trifloxystrobin 50WG at 100ppm after 10 days of incubation (Fig. 1a). However, an inhibition of 21.22 and 25.56 per cent respectively was noticed at 250 and 350ppm on 10th day. Azoxystrobin (Fig. 1b) and kresoxym-methyl 44.3 SC (Fig. 1c) were less toxic to and compatible with *P. indica* at all concentrations tested. Thus all three strobilurin fungicides were compatible with *P. indica*. Pencycuron 22.9% SC (Fig. 1d) also recorded a compatibility of 88.88, 76.66 and 56.67 per cent at 100, 250 and 350 ppm concentration respectively (Fig. 2).

The combination fungicide, trifloxystrobin 25 % + tebuconazole 18.3% SC at 100, 250 and 350 ppm recorded a compatibility of 90.00, 76.67 and 68.89 per cent with *P. indica* (Fig. 3a). On the contrary, azoxystrobin 11% + tebuconazole 18.3% SC (Fig. 3g) and carbendazim 50 WP showed moderate compatibility of 41.11 to 55.56 per cent and 61.11 to 74.44 per cent with the endophyte (Fig. 3b). However triazole fungicides, hexaconazole 5% EC, propiconazole 25%EC, tebuconazole 25.9% EC and difenoconazole 25% EC (Fig. 3c, 3d, 3e, 3f) were highly inhibitory to *P. indica* with cent per cent inhibition. No differences were observed in the nature of mycelial growth when *P. indica* was grown together with compatible fungicides.

Compatibility of fungicides with *P. indica* was tested in poisoned PDB. Mycelial weight of *P. indica* was recorded on 21st day in control as well as treated broth cultures. Fresh mycelial weight of untreated control flasks were 12g and significant reduction in weight was observed with different fungicide concentrations (Fig. 4). Triazole fungicides viz., hexaconazole 5 EC, difenoconazole 25% EC, propiconazole 25% EC and tebuconazole 25.9% EC showed complete inhibition of mycelial growth of *P. indica* at all the concentrations (Table 1). The mycelial growth was significantly higher when grown along with strobilurin fungicides, carbendazim and pencycuron, showing a compatible reaction. Fresh mycelial weight of the endophyte was ranging from 1.1 to 1.6 g for these fungicides at different concentrations. Combination fungicides also depicted a compatible reaction. No differences in nature of mycelial growth were observed in all the fungicides tested; but its varying growth pattern was noticed. 5 to 6 times reduction in weight of mycelia was noticed when it was dried.

3.2 Spore Germination Inhibition Study

Percentage of chlamyospores germinated was calculated after growing *P. indica* with new generation fungicides. Germination of chlamyospores was completely inhibited by four triazole fungicides. However the germination was approximately 60-70 per cent with trifloxystrobin 50 WG, kresoxym-methyl 44.3 SC, azoxystrobin 23SC, pencycuron 22.9% SC and carbendazim 50WP. Combination fungicides also recorded less inhibition in germination of chlamyospores (Fig. 5.).

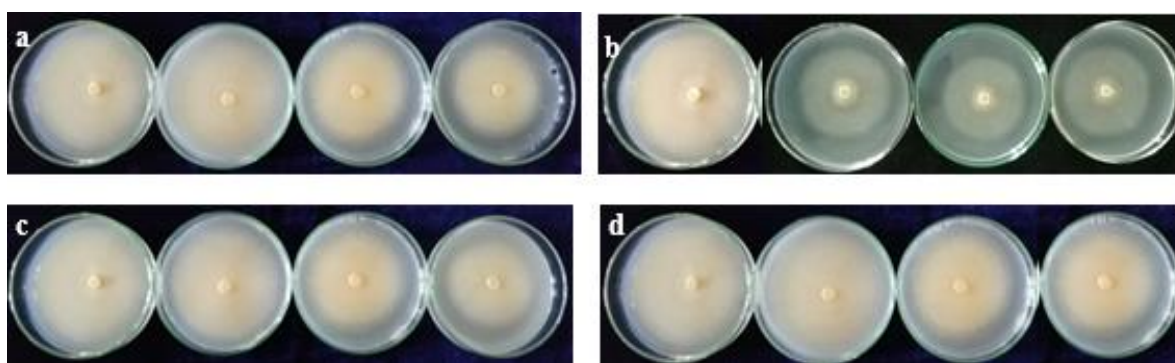


Fig. 1. Compatibility of a) Trifloxystrobin b) Azoxystrobin c) Kresoxym methyl d) Pencycuron with *P. indica* at 100, 250 and 350 ppm concentrations. *P. indica* is grown on PDA amended with respective fungicides and mycelial growth was recorded at various time intervals

Table 1. Mycelial weight of *P. indica* with fungicides *in vitro* in PDB

Treatments (Fungicides)	Mycelial growth (cm) on 10 th day									
	100ppm		250 ppm		350 ppm		500 ppm		1000 ppm	
	F	D	F	D	F	D	F	D	F	D
Trifloxystrobin 50 WG	11±0.86	2.2±0.04	9.46±0.05	1.8±0.12	8.93±0.12	1.78±0.05	4.6±0.04	0.92±0.07	1.73±0.03	0.34±0.02
Kresoxym methyl 44.3 SC	9.1±0.03	1.81±0.05	8.53±0.03	1.7±0.03	6.8±0.01	1.36±0.03	4.13±0.01	0.82±0.03	1.6±0.03	0.32±0.03
Hexaconazole 5 EC	0	0	0	0	0	0	0	0	0	0
Difenoconazole 25% EC	0	0	0	0	0	0	0	0	0	0
Pencycuron 22.9 % SC	10.7±0.18	2.13±0.07	8.53±0.14	1.7±0.02	6.8 ±0.01	1.36±0.02	4.53±0.06	0.9±0.02	1.73±0.01	0.34±0.01
Azoxystrobin 11% +Tebuconazole 18.3 % SC	9.7±0.11	1.7±0.08	8.3±0.04	1.5±0.02	6.2±0.02	0.99±0.07	2.56±0.02	0.51±0.03	0	0
Trifloxystrobin 25%+ Tebuconazole 18.3 % SC	10.8±0.17	2.16±0.08	9.2±0.11	1.8±0.02	8.26±0.01	1.65±0.02	5.2±0.04	1.04±0.02	2.1±0.03	0.56±0.02
Propiconazole 25% EC	0	0	0	0	0	0	0	0	0	0
Bavistin 50WP	8.93±0.08	1.78±0.02	8.13±0.02	1.6±0.01	7.33±0.07	1.46±0.02	5.33±0.14	1.06±0.01	2.67±0.06	0.52±0.03
Azoxystrobin 20% W/V	9.08±0.06	1.81±0.01	8.0±0.02	1.6±0.08	7.32±0.04	1.45±0.02	4.26±0.02	0.85±0.02	2.53±0.03	0.50±0.03
Tebuconazole 25.9% EC	0	0	0	0	0	0	0	0	0	0
Control	12.1±0.22	2.35±0.04	11.9±0.17	2.3±0.03	11.8±0.21	2.35±0.04	11.7±0.17	2.2±0.03	11.8±0.21	2.4±0.02
SEm±	0.000		0.000		0.000		0.010		0.000	
CD(0.05)	0.036		0.013		0.017		0.172		0.008	

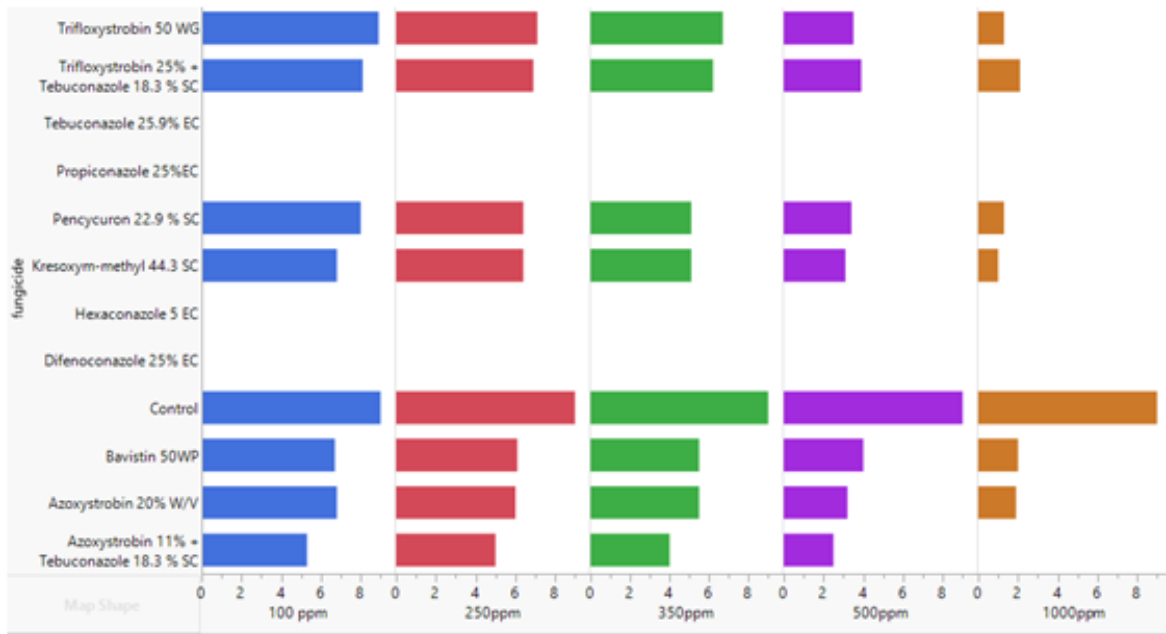


Fig. 2. Mycelial growth *P. indica* at different concentrations 100, 250, 350, 500 and 1000 ppm of fungicides. *P. indica* is grown over PDA amended with respective fungicides and mycelial growth was recorded

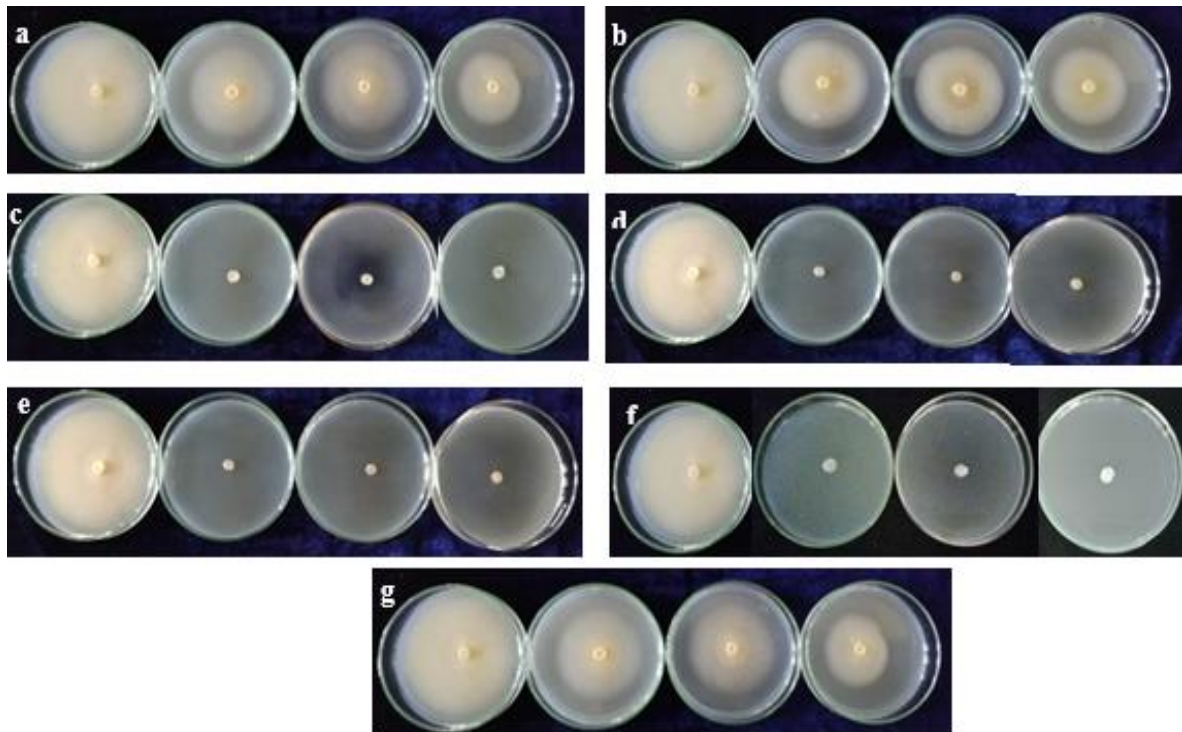


Fig. 3. Compatibility of a) Trifloxystrobin 25 % + Tebuconazole 18.3 % SC b) Carbendazim 50 WP c) Hexaconazole 5 EC d) Propiconazole 25 %EC e) Tebuconazole 25.9% EC f) Difenconazole 25% EC g) Azoxystrobin 11 % + Tebuconazole 18.3 % SC against *P. indica* at 100, 250 and 350 ppm concentrations. *P. indica* is grown over PDA amended with respective fungicides and mycelial growth was recorded



Fig. 4. Compatibility of a) Trifloxystrobin 25 % + Tebuconazole 18.3 % SC b) Pencycuron e) Kresoxym methyl d) Trifloxystrobin e) Azoxystrobin f) Carbendazim 50 WP g) Azoxystrobin 11 % + Tebuconazole 18.3 % SC against *P. indica* at 100, 250 and 350 ppm concentrations. *P. indica* is grown over PDB amended with respective fungicides and mycelial growth is recorded

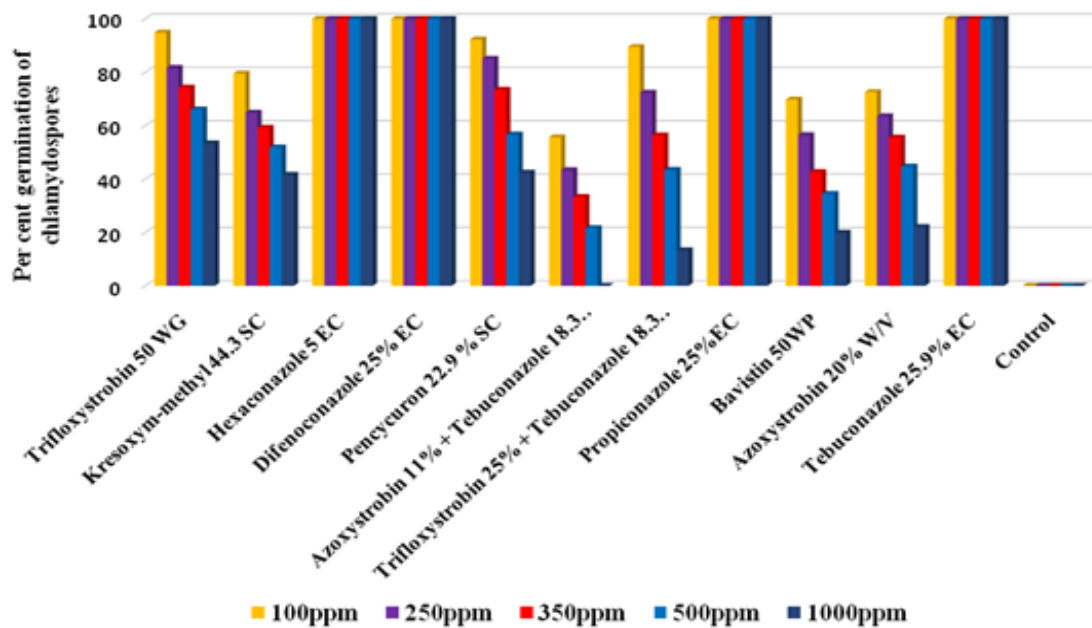


Fig. 5. Effect of fungicides on spore germination of *P. indica*. Percentage of chlamydospores germinated was recorded after allowing the spore suspension to grow against different doses of fungicides

4. DISCUSSION

Use of agrochemicals can never be replaced in agriculture and their impending effects on environment and health should be addressed. Before implementing the usage of endophytic agents in the field, the efficacy of pesticides used in controlling pests and diseases against the particular targeted crop should be determined. It is necessary to understand the compatibility of chemicals with endophytic agents to formulate a better integrated management strategy.

Strobilurins and combination fungicides were most compatible with *P. indica*. The germination of chlamydospores and mycelial growth of *P. indica* were not inhibited by the strobilurins and its combination fungicides. Strobilurin is a natural compound isolated from the mushroom *Strobilurus tenacellus* [28] and contains a toxiphoric (E)- β -methoxyacrylate group [29]. It inhibits the mitochondrial respiration of fungi by binding to the quinol oxidation (Qo) site of cytochrome bc1 [30]. However, the fungicide did not act at the target site of *P. indica*. Similarly, Sendhilvel et al. [19] Anand et al. [20], Archana et al. [21], Louis et al. [31], Suneeta et al. [32], Hanuman and Madhavi [33], Sharma et al. (2017), Widmer [34], Maheswary et al. [35], Sanchez-Montesinos et al. [36] also reported the compatibility of strobilurin with *Trichoderma* spp., a widely used biocontrol agent. Combination fungicide azoxystrobin 18.2 % + difenoconazole 11.4% SC on testing against 5 *Trichoderma* isolates showed 5 to 54 per cent compatibility and 90 to 100 per cent compatibility with tebuconazole 50% + trifloxystrobin 25% WG [37]. To the contrary, Bai et al. [38] could observe cent per cent inhibition with tebuconazole 50% + trifloxystrobin 25% WG and 67.77 per cent with azoxystrobin 18.2% + difenoconazole 11.4% SC.

P. indica was found compatible with pencycuron 22.9% SC at all concentrations (Fig. 1d) with reduced compatibility at very high concentration of 1000 ppm. Pencycuron, a non-systemic phenyl urea fungicide, destroys the microtubules of fungus and changes the osmotic stability and fluidity of the plasma membrane [39] Madhavi et al. [40], Elshahawy et al. [41] Silva et al. [42]; Sanchez-Montesinos et al. [36] also reported a similar compatible action against *Trichoderma* spp.

All four triazole fungicides (Fig 1b and f) completely inhibited the mycelial growth of *P.*

indica and germination of chlamydospores. Triazole fungicides as demethylation inhibitors act on the enzyme C14-demethylase that plays a major role in sterols production, which are essential molecules that ensure stability in the lipid layer [43]. Sterols are considered to be an important component of fungal cell membrane. Similarly, Sarkar et al. [23], Madhusudhanan et al. [44], Madhavi et al. [40], Sreeja and Girija [6,7] Sonavane and Venkataravanappa [45], Maheswary et al. [35] reported complete mycelial inhibition of *Trichoderma* spp. with hexaconazole, difenoconazole and propiconazole as it was found that the fungicide completely inhibits the sterol production of the endophyte. Similarly Bharadwaz et al. [46] also reported complete inhibition or incompatible reaction of *T. viride* with Hexaconazole 5% EC, Propiconazole 25% EC and tebuconazole 25.9% EC. To the contrary Huilgol et al. [47-49] tested the compatibility of *T. harzianum* with four triazole fungicides and observed a mean mycelial growth of 6.84 cm with tebuconazole, 4.08 cm with hexaconazole, 3.71 cm with difenoconazole and 2.02 cm with propiconazole.

5. CONCLUSION

From the present investigation it was found that strobilurin, its combination fungicide and pencycuron are highly compatible with fungal endophyte *P. indica*. Fungicides can be used in *P. indica*-colonized yard long bean plants to protect the plants from anthracnose infection. This method can also reduce the concentration of chemical used and also reduces the buildup of residue on the plant surface thereby reduce health and environmental risks.

ACKNOWLEDGEMENTS

The authors are grateful towards laboratory facilities of Department of Plant Pathology and for funding from Kerala Agricultural University and Jawaharlal Nehru Memorial Fund for Doctoral Studies.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Emechebe AM, Lagoke STO. Recent advances in research on yard long bean diseases. Challenges and opportunities for

- enhancing sustainable yard long bean production. 2002;94-123.
2. Kumar MP. Anthracnose disease of vegetable yard long bean [*Vigna unguiculata* ssp. *sesquipedalis* (L.) Verdcourt]. M. Sc. Thesis, Kerala Agricultural University. 1999; 106.
 3. Sreeja SJ. Integrated management of fusarium wilt and anthracnose of vegetable cowpea (*Vigna unguiculata* subsp. *sesquipedalis* (L.) Verdcourt) using new generation fungicides. Doctoral dissertation. Kerala Agricultural University, Thrissur. 2014;233.
 4. Deeksha J, Tripathi HS. Cultural, biological and chemical control of anthracnose of urdbean. J. Mycol. Plant Pathol. 2002; 32(1):52-55.
 5. Kulkarni SA. Epidemiology and integrated management of anthracnose of green gram. M. Sc. (Agri.) Thesis, University of Agricultural Sciences. 2009;101.
 6. Sreeja SJ, Girija VK. Compatibility of *Trichoderma viride*, *Pseudomonas fluorescens* and *Rhizobium* spp. with selected fungicides. Plant Dis. Res. 2015;30(2):88-189.
 7. Sreeja SJ, Girija VK, Beevi SN. Tebuconazole-A potential and safe new generation fungicide for the management of anthracnose of yard long bean (*Vigna unguiculata* subsp. *sesquipedalis* (L.) Verdcourt). Pesticide Res. J. 2015;27(2):191-198.
 8. Strange RN, Scott PR. Plant disease: A threat to global food security. Ann Rev Phytopathol. 2005;43(1):83-116.
 9. Cook RJ, Baker KF. The Nature and practice of biological control of plant pathogens. American Phytopathol Society. 1983;539.
 10. Lorito M, Woo SL, D'Ambrosio M, Harman GE, Hayes CKCP, Kubicek Scala F. Synergistic interaction between cell wall degrading enzymes and membrane affecting compounds. Mol. Plant Microbe Interact. 1996;9:206-213.
 11. Varma A, Bakshi M, Lou B, Hartmann A, Oelmueller R. *Piriformospora indica*: A novel plant growth-promoting mycorrhizal fungus. Agric. Res. 2012;1(2):117-131.
 12. Waller F, Achatz B, Baltruschat H, Fodor J, Becker K, Fischer M, Heier T, Hüchelhoven R, Neumann C, Von Wettstein D, Franken P. The endophytic fungus *Piriformospora indica* reprograms barley to salt-stress tolerance, disease resistance, and higher yield. Proc. Natl. Acad. Sci. 2005;102(38):13386-13391.
 13. Deshmukh S, Hüchelhoven R, Schäfer P, Imani J, Sharma M, Weiss M, Waller F, Kogel KH. The root endophytic fungus *Piriformospora indica* requires host cell death for proliferation during mutualistic symbiosis with barley. Proc. Natl. Acad. Sci. 2006;103(49):18450-18457.
 14. Kumar M, Yadav V, Tuteja N, Johri AK. Antioxidant enzyme activities in plants colonized with *Piriformospora indica*. Microbiol. 2009;155:780-790.
 15. Yadav V, Kumar M, Deep DK, Kumar H, Sharma R, Tripathi T. A Phosphate transporter from the root endophytic fungus *Piriformospora indica* plays a role in the phosphate transport to the host plant. J. Biol. Chem. 2010;285:26532
 16. Dolatabadi HK, Goltapeh EM, Jaimand K, Rohani N, Varma A. Effects of *Piriformospora indica* and *Sebacina vermifera* on growth and yield of essential oil in fennel (*Foeniculum vulgare*) under greenhouse conditions. J. Basic Microbiol. 2011;51:33-9.
 17. Johnson JM, Alex T, Oelmueller R. *Piriformospora indica*: The versatile and multifunctional root endophytic fungus for enhanced yield and tolerance to biotic and abiotic stress in crop plants. J. Tropical Agric. 2014;52(2):103-122.
 18. Gill SS, Gill R, Trivedi DK, Anjum NA, Sharma KK, Ansari MW, Ansari AA, Johri AK, Prasad R, Pereira E, Varma A. *Piriformospora indica*: Potential and significance in plant stress tolerance. Front. Microbiol. 2016;7:332-352.
 19. Sendhilvel V, Marimuthu T, Raguchander T. Compatibility of azoxystrobin 25 SC with biocontrol agents. Pestoogy. 2004; 28(10):61-64.
 20. Anand T, Prakasam V, Chandrasekaran A, Samiyappan R, Karthikeyan G, Saravanan A. Compatibility of azoxystrobin (Amistar 25 SC) with biocontrol agents. Pestology. 2007;31(5):21-24.
 21. Archana S, Hubballi M, Ranjitham TP, Prabakar K, Raguchander T. Compatibility of azoxystrobin 23 SC with biocontrol agents and insecticides. Madras Agric. J. 2012;99(4-6):374-377.
 22. Bagwan NB. Evaluation of *Trichoderma* compatibility with fungicides, pesticides,

- organic cakes and botanicals for integrated management of soil borne disease of soybean [*Glycine max* (L.) Merrill]. Int. J. Plant Protec. 2010;3(2):206-209.
23. Sarkar S, Narayanan P, Divakaran A, Balamurugan A, Premkumar R. The *In vitro* effect of certain fungicides, insecticides, and biopesticides on mycelial growth in the biocontrol fungus *Trichoderma harzianum*. Turkish J. Biol. 2010;34(4):399-403.
 24. Ranganathswamy M, Patibanda AK, Chandrashekar GS, Sandeep D, Mallesh SB, Kumar HH. Compatibility of *Trichoderma* isolates with selected fungicides *In vitro*. Int. J. Plant Protec. 2012;5(1):12-15.
 25. Johnson JM, Sherameti I, Nongbri PL, Oelmüller R. Standardized conditions to study beneficial and nonbeneficial traits in the *Piriformospora indica* / *Arabidopsis thaliana* interaction. *Piriformospora indica*: Sebaciniales and their Biotechnological Applications. Springer, Berlin, Heidelberg. 2013;325-343.
 26. Vincent JM. Distortion of fungal hyphae in the presence of certain inhibitors. Nature. 1947;159(4051):850-850.
 27. Reshna OP, Beena R, Joy M, Viji MM, Roy S. Elucidating the effect of growth promoting endophytic fungus *Piriformospora indica* for seedling stage salinity tolerance in contrasting rice genotypes. J. Crop Sci. Biotechnol. 2022;25(5):583-598.
 28. Anke T, Oberwinkler F, Steglich W, Schramm G. The strobilurins-new antifungal antibiotics from the basidiomycete *Strobilurus tenacellus* (Pers. ex Fr.) Sing. J. Antibiot. 1977;30:806–810:10.7164
 29. Balba H. Review of strobilurin fungicide chemicals. J. Environ. Sci. Heal. 2007;B 42:441–451.
 30. Feng Y, Huang Y, Zhan H, Bhatt P, Chen S. An overview of strobilurin fungicide degradation: Current status and future perspective. Front. Microbiol. 2020;11:389.
 31. Louis V, Sindu PG, Jiphy Jose P, Pushpalatha PB. Compatibility of *Pseudomonas fluorescens* with fungicides used in Banana cultivation. Int. J. Agric. Innovations Res. 2016;5(3):487-488.
 32. Suneeta P, Kumar SV, Aiyathan KEA, Nakkeeran S. Promissory Action of *Trichoderma* spp. and fungicides in the management of Fusarium wilt of gerbera. J. Pure Appl. Microbiol. 2017;11(1):241-247.
 33. Hanuman LN, Madhavi GB. Compatibility of *Pseudomonas fluorescens* with pesticides *In vitro* Int. J. Curr.Microbiol. App. Sci. 2018;7(03):1-7.
 34. Widmer TL. Compatibility of *Trichoderma asperellum* isolates to selected soil fungicides. Crop Protec. 2019;120:91-96.
 35. Maheshwary N, Gangadhara Naik B, Amogha varsha Chittaragi M, Naik SK, Nandish M. Compatibility of *Trichoderma asperellum* with fungicides. Pharma Innov. J. 2020;9:136-140.
 36. Sanchez-Montesinos B, Santos M, Moreno-Gavira A, Marín-Rodulfo T, Gea FJ, Dianez F. Biological control of fungal diseases by *Trichoderma aggressivum* f. sp. *europaeum* and its compatibility with fungicides. J. Fungi. 2021;7(8):598-616.
 37. Dinkwar GT, Yadav VK, Kumar A, Nema S, Mishra S. Compatibility of Fungicides with Potent *Trichoderma* Isolates. Int. J. Plant Soil Scie. 2023;35(18):1934-1948.
 38. Bai AT, Vibha JB, Nair R, Upadhyay A. Compatibility of *Trichoderma viride* with commonly used fungicides in management of early blight pathogen of tomato. The Pharma Inno. J. 2022;11:1734-1736.
 39. Ueyama I, Kurahashi Y. Pencycuron, a Phenylurea Fungicide for *Rhizoctonia solani* in Modern Crop Protection Compounds. W. Kramer and U. Schirmer (eds.). 2007; 2:591-604.
 40. Madhavi GB, Bhattiprolu SL, Reddy VB. Compatibility of biocontrol agent *Trichoderma viride* with various pesticides. J. Hortic. Scie. 2011;6(1):71 -73.
 41. Elshahawy IE, Haggag KH, Abd-EI-Khair H. Compatibility of *Trichoderma* spp. with seven chemical fungicides used in the control of soil borne plant pathogens. Res. J. Pharm. Biol. Chem. Sci. 2016;7(1):1772-1785.
 42. Silva MAFD, Moura KED, Moura KED, Salomão D, Patricio FRA. Compatibility of *Trichoderma* isolates with pesticides used in lettuce crop. Summa Phytopathologica. 2018;44:137-142.
 43. Dupont S, Beney L, Ferreira T, Gervais P. Nature of sterols affects plasma membrane behavior and yeast survival during dehydration. Biochim Biophys Acta Biomembr. 2011;1808:1520–1528.

44. Madhusudhanan P, Gopal K, Haritha V, Sangale UR, Rao SVRK. Compatibility of *Trichoderma viride* with fungicides and efficiency against *Fusarium solani*. J. Plant Dis. Sci. 2010;5(1):23-26.
45. Sonavane P, Venkataravanappa V. Compatibility Studies of *Trichoderma harzianum* isolate with fungicides used against soil borne disease in Coorg Mandarin-Pepper-Coffee Plantations. Int. J. Curr. Microbiol. App. Sci. 2017;6(8):346-354.
46. Bharadwaz P, Nath BC, Chetia R, Saikia S, Bora P, Bhattacharyya PN. *In-vitro* studies on the compatibility of *Trichoderma viride* with commonly used agrochemicals in the vegetable cropping system. Pest Manag. Hortic.Ecosyst. 2023;29(1):136-143.
47. Huilgol SN, Pratibha MP, Hegde GM, Banu H. Evaluation, and compatibility of new fungicides with *Trichoderma harzianum* for managing the charcoal rot of soybean. Pharma Innov. J. 2022;11:659-664.
48. Anand T, Chandrasekaran A, Kuttalam S, Senthilraja G, Samiyappan R. Integrated control of fruit rot and powdery mildew of chilli using the biocontrol agent *Pseudomonas fluorescens* and a chemical fungicide. Biol. Control. 2010;52(1):1-7.
49. Anand Sharma D, Singh RP. Compatibility of different fungicides with *Trichoderma harzianum* rifai strain PBAT-21. Pesticide Res. J. 2017;29(1):42-47.

© Copyright (2024): Author(s). The licensee is the journal publisher. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:

The peer review history for this paper can be accessed here:

<https://www.sdiarticle5.com/review-history/116370>